Asymmetric Synthesis and Antiviral Activities of L-Carbocyclic 2',3'-Didehydro-2',3'-dideoxy and 2',3'-Dideoxy Nucleosides

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Received March 22, 1999

Asymmetric syntheses of L-carbocyclic 2',3'-didehydro-2',3'-dideoxy- and 2',3'-dideoxypyrimidine and purine nucleoside analogues were accomplished, and their anti-HIV and anti-HBV activities were evaluated. The key intermediate, (1S, 4R)-1-benzoyloxy-4-(*tert*-butoxymethyl)cyclopent-2-ene (7), was prepared by benzovlation of the alcohol 2, selective deprotection of the isopropylidene group of **3**, followed by thermal elimination via cyclic ortho ester or deoxygenation via cyclic thionocarbonate. The target compounds were also synthesized by thermal elimination via cyclic ortho esters from protected nucleosides. It was found that L-carbocyclic 2',3'-didehydro-2',3'-dideoxyadenosine (34) exhibited potent anti-HBV activity (EC₅₀ = 0.9 μ M) and moderate anti-HIV activity (EC₅₀ = 2.4 μ M) in vitro without cytotoxicity up to 100 μ M.

Introduction

Since the discovery of the natural products (-)aristeromycin and neplanocin A as biologically interesting carbocyclic nucleosides, a number of carbocyclic nucleosides have been synthesized and have shown interesting antitumor¹ and antiviral activity against herpes virus,² cytomegalovirus,³ hepatitis B virus,⁴ and human immunodeficiency virus (HIV).⁵ These compounds possess greater metabolic stability to nucleoside phosphorylases, which cleave the glycosidic bond of normal nucleosides.⁶ Among them, carbovir⁷ and its 6-cyclopropylamino analogue 1592U89 (abacavir)⁸ are the most interesting compounds as they both exhibit potent anti-HIV activity. Abacavir has been reported to reduce the viral load of greater than 99% with significant improvement in CD4 counts, and it has recently been approved by the FDA. More recently, a novel carbocyclic nucleoside, BMS-200475, has been reported to have potent anti-hepatitis B virus activity and is currently undergoing phase II clinical trials.⁹

Recently, a number of nucleosides with the unnatural L-configuration have emerged as potent antiviral agents against HIV and HBV, which include (-)-(2'R,5'S)-1-(2-hydroxymethyloxathiolan-5-yl)cytosine (3TC),^{10,11} (-)- β -L-2',3'-dideoxy-5-fluoro-3'-thiacytidine (FTC),¹² β -L-2',3'-dideoxy-5-fluorocytidine (L-FddC),¹³ and β -L-2'fluoro-5-methylarabinofuranosyl uracil (L-FMAU).¹⁴ Both 3TC and FTC were found to have more potent antiviral activity against HIV and HBV than their D-counterparts with reduced toxicity.^{15,16} Two other L-nucleosides, β -L-

d4C and β -L-Fd4C, were reported to exhibit potent anti-HIV and anti-HBV activities.¹⁷ The synthesis of the enantiomers of abacavir was reported¹⁸ and the Denantiomer of abacavir also exhibited anti-HBV activity.¹⁹ In connection with L-nucleosides, we have reported the synthesis and antiviral activity of β -L-2',3'-didehydro-2',3'-dideoxy purine nucleoside analogues, among which the adenosine derivative (β -L-d4A) exhibited significant anti-HIV and anti-HBV activities.²⁰ The racemic mixtures of some carbocyclic 2',3'-didehydro-2',3'-dideoxy purine derivatives were reported to exhibit anti-HIV activity.²¹ Carbovir was the first carbocyclic nucleoside analogue which exhibits potent anti-HIV activity in vitro. The first preparation of the D- and Lisomers of carbovir was reported by Vince et al.²² The L-carbovir was prepared either by chemoenzymatic syntheses, 2^{2-24} or by a stereoselective method via [2+3] asymmetric cycloaddition.²⁵ The anti-HIV and anti-HBV activities of carbovir reside in its natural β -D-enantiomer.²⁶ It was reported that the triphosphates of β -D- and β -L-carbovir are approximately equipotent as HIV reverse transcriptase inhibitors²⁴ and the β -L-carbovir triphosphate exhibited more potent anti-HBV activity.²⁷

These findings prompted us to synthesize L-carbocyclic nucleosides in the search for new antiviral agents. Recently, we have reported a preliminary result of carbocyclic β -L-2',3'-didehydro-2',3'-dideoxy adenine as an anti-HIV agent.²⁸ As part of our ongoing drug discovery efforts, herein we report the full account of the asymmetric syntheses of carbocyclic β -L-2',3'-didehydro-2',3'-dideoxy and β -L-2',3'-dideoxy pyrimidine and purine nucleoside analogues together with their anti-HIV and anti-HBV activities.

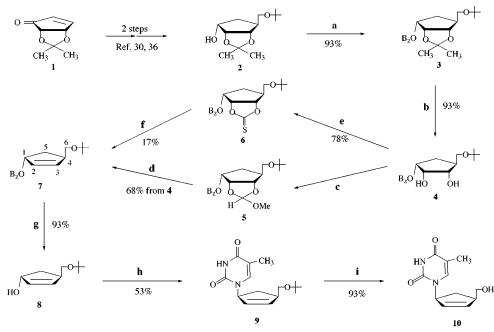
For the synthesis of L-carbocyclic nucleosides, we utilized the reported (+)-cyclopentenone 1 as a chiral starting material, which was prepared in three steps from D-ribose.²⁹ The alcohol **2** was previously reported by our group from the regioselective addition of tertbutoxymethyl group to the enone 1, followed by DIBAL-H

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Scheme 1^a



^{*a*} Reagents and conditions: (a) BzCl, Pyr, rt, 12 h; (b) concentrated HCl:MeOH (1:70, v/v), rt, 2.5 h; (c) CH(OMe)₃, pyridinium *p*-toluenesulfonate, rt, 2 h; (d) Ac₂O, 120–130 °C, 6 h; (e) 1,1'-thiocarbonyldiimidazole, MeCN, rt, 24 h; (f) 1,3-dimethyl-2-phenyl-1,3,2diazaphospholidine, THF, rt, 20 h; (g) 2 N NaOH/MeOH, rt, 1.5 h; (h) (1) N^3 -benzoylthymine, Ph₃P, diethyl azodicarboxylate, dioxane, rt, 6 h; (2) NH₃/MeOH, rt; (i) CF₃CO₂H/H₂O (2:1), 50 °C, 3 h.

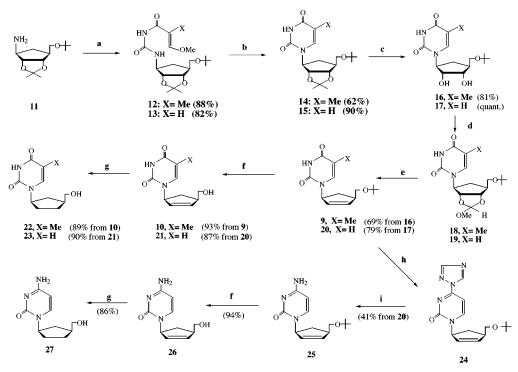
reduction (Scheme 1).³⁰ Treatment of compound **2** with benzoyl chloride in pyridine gave the benzoate 3 in 93% yield. Selective deprotection of the isopropylidene protection group of compound 3 with a mixture of concentrated HCl and methanol (1: 70, v/v) at room temperature for 2.5 h gave the diol 4 in 93% yield. The diol 4 was treated with trimethyl orthoformate at room temperature for 2 h in the presence of catalytic amounts of pyridinium *p*-toluene-sulfonate to afford the cyclic ortho ester 5, which was subjected to a thermal elimination reaction³¹ with acetic anhydride at 120-130 °C for 6 h to give the required cyclopentene 7. Another method of deoxygenation of the diol 4 to 7 via cyclic thionocarbonates was also explored. The yield was, however, lower than that of the above method as briefly described below: Compound 4 was treated with 1,1'-(thiocarbonyl)diimidazole at room temperature to give cyclic thionocarbonate 6, which was deoxygenated to 7 under mild conditions using 1,3-dimethyl-2-phenyl-1,3,2-diazaphospholidine.32

For structural identification, NOESY experiments were performed, which indicated that there is a correlation between H-1 and H-6 of compound **7**, suggesting that H-1 and H-6 are on the same side. The benzoyl group of compound **7** was removed with 2 N NaOH in methanol to give the alcohol **8**, which was then reacted with a heterocyclic base under Mitsunobu conditions.^{33,34} Thymidine analogue was prepared by the reaction of **8** with N^3 -benzoylthymine³⁵ in the presence of triphenylphosphine and DEAD at room temperature to give **9** in 53% yield. Deblocking of compounds **9** with trifluoroacetic acid/H₂O (2:1) at 50 °C for 3 h provided thymine analogue **10** in 93% yield.

Other carbocyclic L-2',3'-didehydro-2',3'-dideoxy and 2',3'-dideoxy pyrimidine and purine nucleosides were also prepared by thermal elimination reactions via cyclic ortho esters from protected nucleosides (Scheme 2 and

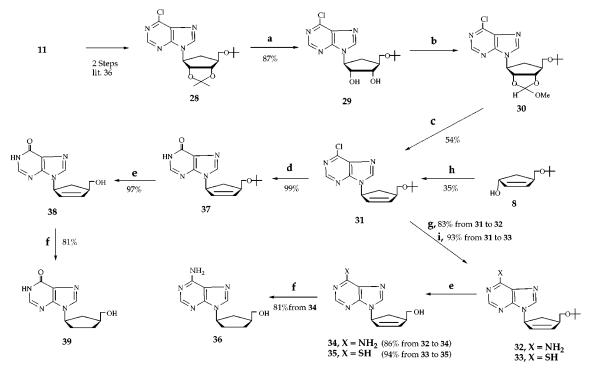
Scheme 3). Treatment of $11^{30,36}$ with β -methoxy- α methacryloyl isocyanate and β -methoxy acryloyl isocyanate using the methodology reported by Shealy et al.^{37,38} gave acrylurea **12** (X = Me) and **13** (X = H), respectively. Reaction of 12 and 13 with 30% NH₄OH and ethanol at 80-100 °C in a steel bomb gave the corresponding thymine 14 and uracil 15 derivatives, respectively. Treatment of compound 14 and 15 with a mixture of concentrated HCl and methanol (1: 65, v/v) at room temperature for 2.5 h gave the diol 16 and 17 in 81% and quantitative yields, respectively. Ortho esterification of compound 16 and 17 with trimethyl orthoformate in the presence of pyridinium *p*-toluenesulfonate gave the cyclic ortho esters 18 and 19, which were then heated with acetic anhydride at 120-130 °C to afford 9 and 20, respectively. Deprotection of compounds 9 and 20 employing the conditions described above provided 10 (93% yield) and 21 (87% yield), respectively. Preparation of the cytosine derivative 26 was accomplished by the reported method.³⁹ The treatment of 20 with 4-chlorophenyl dichlorophosphate and 1,2,4-triazole in pyridine gave the triazole derivative **24**. Subsequent hydrolysis of 24 with concentrated NH₄OH gave 25 (41% yield), which was deblocked with trifluoroacetic acid/H₂O to afford the cytosine nucleoside 26 in 94% yield.

The 6-chloropurine derivative **28** was prepared in two steps from **11** (Scheme 3).³⁶ The isopropylidene protection group of compound **28** was removed using a mixture of HCl and methanol (1: 70, v/v) to give compound **29** in 87% yield. Treatment of the diol **29** with trimethyl orthoformate in the presence of pyridinium *p*-toluenesulfonate gave **30**, which was then subjected to a thermal elimination reaction with acetic anhydride to give compound **31**. The 6-chloropurine nucleoside analogue **31** was also obtained in 35% yield by the reaction of alcohol **8** with 6-chloropurine in the presence of Scheme 2^a



^{*a*} Reagents and conditions: (a) β-methoxy-α-methacryloyl isocyanate, DMF, -20 to 20 °C, 10 h (for **12**) or β-methoxyacryloyl isocyanate, DMF, -20 to 20 °C, 10 h (for **13**); (b) 30% NH₄OH, EtOH or/and dioxane, 80–100 °C, 10–24 h; (c) concentrated HCl:MeOH (1:65 to 70, v/v), rt, 2.5–3 h; (d) CH(OMe)₃, pyridinium *p*-toluene-sulfonate; rt, 30 min; (e) Ac₂O, 120–130 °C, 8 h; (f) CF₃CO₂H/H₂O (2:1), 50 °C, 3 h; (g) 10% Pd/C, MeOH, 15 psi; (h) *p*-chlorophenylphosphorodichloridate, 1,2,4-triazole, Py, rt, 24 h; (i) 30% NH₄OH, dioxane, rt, 24 h.

Scheme 3^a

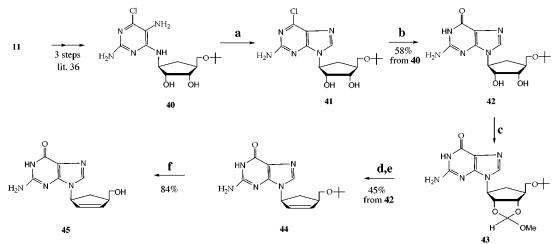


^{*a*} Reagents and conditions: (a) concentrated HCl:MeOH (1:70, v/v), rt, 3 h; (b) CH(OMe)₃, pyridinium *p*-toluene-sulfonate, rt, 2 h; (c) Ac₂O, 120–130 °C, 12 h; (d) HSCH₂CH₂OH, MeOH, reflux, 24 h; (e) CF₃CO₂H/H₂O (2:1), 50 °C, 3 h; (f) 10% Pd/C, MeOH, rt, 15 psi, 3 h; (g) NH₃/MeOH, 80–90 °C, 20 h; (h) 6-chloropurine, Ph₃P, diethyl azodicarboxylate, dioxane, rt, 10 h; (i) thiourea, EtOH, reflux, 1 h.

triphenylphosphine and diethyl azodicarboxylate (DEAD) at room temperature (Scheme 3). Treatment of the compound **31** with saturated ammonia in methanol in a steel bomb at 80 to 90 °C provided the adenine derivative **32** in 83%. The *tert*-butyl group of **32** was removed

by CF₃CO₂H/H₂O (2:1, v/v) at 50 °C to afford β -L-2',3'didehydro-2',3'-dideoxyadenosine **34** in 86% yield. The 6-mercaptopurine analogue **35** was obtained by the reaction of **31** with thiourea in refluxing ethanol followed by deprotection with a mixture of CF₃CO₂H and





^{*a*} Reagents and conditions: (a) CH(OMe)₃, HCl, DMF, $0-5 \degree C$, 8 h then rt 18 h; (b) HSCH₂CH₂OH, MeOH, reflux, 18 h; (c) CH(OMe)₃, pyridinium *p*-toluene-sulfonate, rt, 1 h; (d) Ac₂O, 120–130 \degree C, , 12 h; (e) 30% NH₄OH, MeOH, H₂O, 65 °C, 1.25 h; (e) CF₃CO₂H/H₂O (2:1), 50 °C, 3 h.

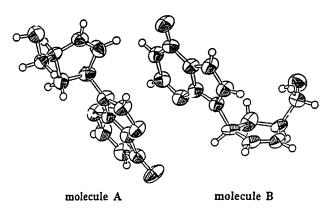


Figure 1. ORTEP drawing of compound 38 (C-L-d4I).

H₂O. Treatment of compound **31** with mercaptoethanol and sodium methoxide in refluxing methanol followed by deblocking with CF₃CO₂H/H₂O gave carbocyclic β -L-2',3'-didehydro-2',3'-dideoxyinosine (**38**). 2',3'-Saturated carbocyclic pyrimidine (**22, 23,** and **27**) and purine (**36, 39**) nucleosides were prepared by the catalytic hydrogenation of the corresponding 2',3'-unsaturated nucleosides with palladium on activated carbon (Schemes 2 and 3).

For the synthesis of L-carbovir **45**, we utilized 6-chloro-2,5-diaminopyrimidine analogue **40** as an intermediate, which was synthesized from cyclopentylamine **11** in three steps (Scheme 4).^{30,36} Treatment of **40** with triethyl orthoformate in the presence of concentrated HCl gave the 2-amino-6-chloropurine analogue **41**, which was treated with a mixture of mercaptoethanol and sodium methoxide in methanol to give the guanine derivative **42**. Conversion of **42** to L-carbovir **45** was accomplished via a thermal elimination reaction with acetic anhydride described as above (Scheme 4).

The X-ray structure of the inosine derivative **38**⁴⁰ contains two molecules (**A** and **B**) with different conformational features (Figure 1). One molecule (molecule **A**) adopts a sugar puckering with the pseudorotation phase angle $P = 264.4^{\circ}$ and $\nu_{max} = 23.60^{\circ}$, the 5'-OH orientation $\gamma = 68.1^{\circ}$ (C_{3'}-C_{4'}-C_{6'}-O_{6'}), and the base orientation, $\chi = 98.9^{\circ}$ (C_{5'}-C_{1'}-N₉-C₄). The other (molecule **B**) has the C5'-exo conformation with $P = 80.8^{\circ}$

and $\nu_{\text{max}} = 9.97^{\circ}$, the 5'-OH orientation $\gamma = -171.6^{\circ}$ (C_{3'}-C_{4'}-C_{6'}-O_{6'}), and the base orientation, $\chi = 68.5^{\circ}$ (C_{5'}-C_{1'}-N₉-C₄) (Table 1).

The anti-HIV and anti-hepatitis B virus (HBV) activities of synthesized L-carbocyclic nucleosides were evaluated in peripheral blood mononuclear (PBM) and 2.2.15 cells, respectively (Table 2). It was found that β -Lcarbocyclic 2',3'-didehydro-2',3'-dideoxyadenosine (34) exhibited potent anti-HBV activity (EC₅₀ = $0.9 \ \mu M$ in 2.2.15 cells) and moderately potent anti-HIV activity $(EC_{50} = 2.4 \ \mu M, EC_{90} = 11.7 \ \mu M$ in PBM cells) without cytotoxicity up to 100 μ M in PBM, CEM, and Vero cells. The cytosine analogue 26 displayed some anti-HBV activity (EC₅₀ = 10.0 μ M in 2.2.15 cells) and anti-HIV activity (EC₅₀ = 35.8 μ M, EC₉₀ = 65.9 μ M in PBM cells). In the carbocyclic L-d2N series, only adenine analogue **36** exhibited weak anti-HIV activity with an EC₅₀ value of 27 μ M. The thymine derivative **10** did not display any antiviral activity, but showed weak cytotoxicity.

Experimental Section

Melting points were determined on a Mel-temp II apparatus and are uncorrected. Nuclear magnetic resonance spectra were recorded on a Bruker 400 AMX spectrometer at 400 MHz for ¹H NMR and 100 MHz for ¹³C NMR with tetramethylsilane as the internal standard. Chemical shifts (δ) are reported in parts per million (ppm), and signals are reported as s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet), or br s (broad singlet). UV spectra were recorded on a Beckman DU-650 spectrophotometer. Optical rotations were measured on a Jasco DIP-370 digital polarimeter. Mass spectra were recorded on a Micromass Autospec high-resolution mass spectrometer. TLC was performed on Uniplates (silica gel) purchased from Analtech Co. Column chromatography was performed using silica gel G (TLC grade, >440 mesh) for vacuum flash column chromatography. Elemental analyses were performed by Atlantic Microlab Inc., Norcross, GA.

(1*R*,2*R*,3*S*,4*S*)-1-Benzoyloxy-4-(*tert*-butoxymethyl)-2,3-(isopropylidenedioxy)cyclopentane (3). Benzoyl chloride (0.28 mL, 2.4 mmol) was added dropwise to a solution of alcohol 2 (0.2 g, 0.81 mmol) in pyridine (2 mL) at 0 °C. The reaction mixture was stirred at 0 °C for 2 h at room temperature for 12 h. After the reaction mixture was quenched by adding ice, it was stirred for 15 min and then the solvent was removed in vacuo. The residue was treated with ethyl acetate (15 mL) and water (5 mL). The organic layer was washed with saturated NaHCO₃ (5 mL) and brine (5 mL) and dried

Table 1. Co	onformational	Parameters	for	Compound	38
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pseudorotation		torsion an	ngles (deg)		
compd 38	angle, P (deg)	$C_{3'-}C_{4'-}C_{6'-}O_{6'}(\gamma)$	$C_{5'-}C_{1'-}N_{9'-}C_4(\chi)$	conformation	
Α	264.4	68.1	98.9	C5'-endo	
В	80.8	-171.6	68.5	C5'-exo	

Table 2.	Anti-HIV	and Anti-HBV	Activity a	and Cytotoxicitie	es of L-Carboc	yclic d4-	and d2-Nucleosides ^a
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	anti-HIV (μ M) in PBM cells		anti-HBV (μ M) in 2.2.15 cells	cytotoxicities (IC ₅₀ , μ M)		
no.	EC ₅₀	EC ₉₀	EC ₅₀	PBM	CEM	Vero
10	>100	-	>10	66.6	>100	49.4
(C-L-d4T) 21	>100	-	>10	>100	>100	>100
(C-L-d4U) 22 (C-L-d2T)	>100	-	>10	>100	>100	171
(C-L-d2U)	>100	-	>10	>100	>100	>100
26	35.8	65.9	10.0	>100	>100	>100
(C-L-d4C) 30 (C-L-d2C)	>100	-	>10	>100	>100	>100
34 (C-L-d4A)	2.4	11.7	0.9	>100	>100	>100
35 (C-L-d4MP)	>100	-	>10	>100	>100	>100
36 (C-L-d2A)	27.0	-	>50	>100	>100	>100
38 (C-L-d4I)	>100	-	>10	>100	>100	>100
39 (C-L-d2I)	>100	-	>10	>100	>100	>100
45 L-carbovir	>200 ²²	-	>200 ²⁶	-	-	-

^{*a*} -: not determined.

(MgSO₄). After filtration and evaporation, the residue was purified by silica gel column chromatography with 1-3% EtOAc in hexanes to give **3** (265 mg, 93.4%) as a syrup: $[\alpha]^{25}_{\rm D}$ 55.61° (*c* 0.82, CHCl₃); ¹H NMR (CDCl₃) δ 7.41–8.10 (m, 5H, Ar–H), 5.28 (m, 1H, 1-H), 4.77 (t, J = 5.6 Hz, 1H, 2-H), 4.50 (d, J = 5.5 Hz, 1H, 3-H), 3.38 (dd, J = 4.3, 8.7 Hz, 1H, 6-Ha), 3.31 (dd, J = 4.3, 8.7 Hz, 1H, 6-Hb), 2.26–2.37 (m, 2H, 5-H), 1.98 (m, 1H, 4-H), 1.41 (s, 3H, CH₃), 1.31 (s, 3H, CH₃), 1.19 (s, 9H, *tert*-butyl); HR-FAB MS Obsd, *m*/*z* 349.1998; Calcd for C₂₀H₂₈O₅, *m*/*z* 349.2015 (M + H)⁺. Anal. (C₂₀H₂₈O₅) C, H.

(1R,2R,3S,4S)-1-Benzoyloxy-2,3-dihydroxy-4-(tert-butoxymethyl)cyclopentane (4). A solution of compound 3 (3.15 g, 9.04 mmol) in a mixture of concentrated HCl and MeOH (324 mL, concentrated. HCl: MeOH = 1:70, v/v) was stirred at room temperature for 2.5 h. The reaction mixture was cooled in an ice bath, neutralized with NaHCO₃ solid, and filtered. The filtrate was concentrated to dryness, and the residue was purified by silica gel column chromatography with 10-25% EtOAc in hexanes to give 4 (2.60 g, 93.3%) as a syrup: $[\alpha]^{25}_{D}$ –28.61° (*c* 0.87, CHCl₃); ¹H NMR (DMSO-*d*₆) δ 7.49–8.04 (m, 5H, Ar–H), 5.04 (m, 1H, 1-H), 4.74 (d, J = 5.1Hz, 1H, OH, D₂O exchangeable), 4.52 (d, J = 6.5 Hz, 1H, OH, D_2O exchangeable), 3.97 (dd, J = 4.6, 9.2 Hz, 1H, 2-H), 3.70 (dd, J = 6.7, 11.3 Hz, 1H, 3-H), 3.14–3.37 (m, 2H, 6-H), 2.17 (m, 1H, 5-Ha), 1.77-1.95 (m, 2H, 4-H and 5-Hb), 1.13 (s, 9H, tert-butyl); HR-FAB MS Obsd, m/z 309.1711; Calcd for C₁₇H₂₅O₅, m/z 309.1702 (M + H)⁺. Anal. (C₁₇H₂₄O₅) C, N.

(1a*S*,3a*S*,4*R*,6*S*)-4-Benzoyloxy-6-(*tert*-butoxymethyl)-3a,4,5,6,6a-tetrahydrocyclopenta-1,3-dioxole-2-thione (6). 1,1'-Thiocarbonyldiimidazole (295 mg, 1.61 mmol) was added to a solution of compound 4 (0.2 g, 0.649 mmol) in dry MeCN (15 mL). The mixture was stirred under nitrogen at room temperature for 24 h. The solvent was removed in a vacuum, and the residue was purified by silica gel column chromatography with 1–3% EtOAc in hexanes to give **6** (177 mg, 78%) as a solid: mp 68–70 °C; $[\alpha]^{28}_D$ 122.35° (*c* 0.27, CHCl₃); ¹H NMR (CDCl₃) δ 7.43–8.08 (m, 5H, Ar–H), 5.61 (m, 1H, 1-H), 5.39 (t, J = 6.1 Hz, 1H, 2-H), 5.17 (d, J = 6.5 Hz, 1H, 3-H), 3.55 (dd, J = 3.4, 8.9 Hz, 1H, 6-Ha), 3.37 (dd, J = 3.1, 8.9 Hz, 1H, 6-Hb), 2.73 (m, 1H, 5-Ha), 2.16–2.29 (m, 2H, 4-H and 5-Hb), 1.19 (s, 9H, *tert*-butyl); ¹³C NMR (CDCl₃) δ 191.51, 165.88, 133.41, 129.94, 128.46, 89.40, 84.25, 74.05, 73.55, 62.65, 42.56, 32.11, 27.35; HR-FAB MS Obsd, *m*/*z* 351.1251; Calcd for C₁₈H₂₃O₅S, *m*/*z* 351.1266 (M + H)⁺. Anal. (C₁₈H₂₂O₅S) C, H, S.

(1*S*,4*R*)-1-Benzoyloxy-4-(*tert*-butoxymethyl)cyclopent-2-ene (7). Method a. A suspension of compound 4 (1.69 g, 5.48 mmol) and pyridinium *p*-toluene-sulfonate (1.54 g, 6.0 mmol) in trimethyl orthoformate (12 mL) was stirred at room temperature for 2 h. The reaction mixture was diluted with ether (160 mL) and washed with aqueous NaHCO₃ (2 × 40 mL) and brine (40 mL). The organic layer was dried (MgSO₄), filtered, and concentrated to dryness. The residue, after being coevaporated with benzene (2 × 10 mL), was treated with acetic anhydride (17 mL). The reaction mixture was heated at 120–130 °C for 6 h, and the mixture was concentrated to dryness. The residue gel column chromatography with 1–2% EtOAc in pentane to 7 (1.023 g, 68% from 4).

Method b. A solution of compound **6** (148 mg, 0.42 mmol) in dry THF was treated with 1,3-dimethyl-2-phenyl-1,3,2-diazaphospholidine (120 mg, 0.6 mmol) and then stirred at room temperature for 20 h. The reaction mixture was concentrated to dryness, and the residue was purified by silica gel column chromatography with 1% Et₂O in hexanes to give **7** (19.4 mg, 17%): $[\alpha]^{27}_{D}$ 259.89° (*c* 1.45, CHCl₃); ¹H NMR (CDCl₃) δ 7.40–8.03 (m, 5H, Ar–H), 6.15 (m, 1H, 2-H), 5.98 (m, 1H, 3-H), 5.95 (m, 1H, 1-H), 3.28 (d, *J* = 6.9 Hz, 1H, 6-H), 3.13 (m, 1H, 4-H), 2.10 (m, 1H, 5-Hab), 1.19 (s, 9H, *tert*-butyl); ¹³C NMR (CDCl₃) δ 166.59, 140.05, 132.76, 130.65, 130.07, 129.60, 128.27, 80.66, 72.69, 65.50, 45.68, 34.35, 27.52; HR-FAB MS Obsd, *m*/*z* 275.1645; Calcd for C₁₇H₂₃O₃, *m*/*z* 275.1647 (M + H)⁺. Anal. (C₁₇H₂₂O₃) C, H.

(1*S*,4*R*)-4-(*tert*-Butoxymethyl)-1-hydroxycyclopent-2ene (8). Compound 7 (578 mg, 2.1 mmol) was treated with a mixture of MeOH (35 mL) and 2 N NaOH (4.5 mL). The reaction mixture was stirred at room temperature for 1.5 h and then neutralized with dry ice. The mixture was concentrated in a vacuum to dryness, and the residue was treated with EtOAc (75 mL) and washed with aqueous NaHCO₃ (2 \times 10 mL) and brine (15 mL). The organic layer was dried (MgSO₄), filtered, and concentrated to give crude **8** (335 mg, 93.4%) as a syrup, which was used for the next step without further purification.

(1'*S*,4'*R*)-1-[4-(*tert*-Butoxymethyl)cyclopent-2-enyl]thymine (9). Method a. A solution of diethyl azodicarboxylate (DEAD) (160 mg, 0.92 mmol) in dioxane (3 mL) was slowly added to a suspension of compound **8** (78 mg, 0.46 mmol), triphenylphosphine (240 mg, 0.92 mmol), and N^3 -benzoylthymine (211 mg, 0.92 mmol) in dry dioxane (4.5 mL). The mixture was stirred at room temperature for 6 h. The solvent was removed under reduced pressure, and the residue was treated with saturated ammonia in MeOH (5 mL). The mixture was stirred at room temperature for 10 h and then concentrated to dryness under reduced pressure. The residue was purified by preparative TLC with 1% MeOH in CHCl₃ to give **9** (68 mg, 53.4%) as a white solid.

Method b. A suspension of compound 16 (119 mg, 0.38 mmol) and pyridinium p-toluene-sulfonate (105 mg, 0.42 mmol) in trimethyl orthoformate (3 mL) was stirred at room temperature for 30 min. The reaction mixture was diluted with EtOAc (20 mL) and washed with saturated NaHCO₃ (2 \times 5 mL) and brine (10 mL). The organic layer was dried (MgSO₄), filtered, and concentrated. The residue was coevaporated with benzene (2×5 mL) and treated with Ac₂O (2 mL). The mixture was heated at 120-130 °C for 8 h and concentrated to dryness. The residue was treated with saturated ammonia in MeOH (8 mL), and the mixture was stirred at room temperature for 8 h. The residue was purified by silica gel column chromatography with 2-5% EtOAc in CHCl₃ to give 9 (73 mg, 68.8%) as a solid: mp 122–124 °C; UV (MeOH) λ_{max} 272 nm; $[\alpha]^{24}$ 59.38° (c 0.33, CHCl₃); ¹H NMR (CDCl₃) δ 8.32 (br s, 1H, NH, D₂O exchangeable, 2-H), 7.22 (s, 1H, 6-H), 6.08 (m, 1H, 2'-H), 5.72 (m, 1H, 1'-H), 5.59 (m, 1H, 3'-H), 3.50 (dd, J = 8.9, 4.1 Hz, 1H, 6'-Ha), 3.34 (dd, J = 8.9, 4.6 Hz, 1H, 6'-Hb), 2.95 (m, 1H, 4'-H), 2.66 (dt, J = 14.0, 9.1, 9.1 Hz, 1H, 5'-Ha), 1.90 (s, 3H, 5-CH₃), 1.49 (m, 1H, 5'-Hb), 1.18 (s, 9H, tert-butyl); HR-FAB MS Obsd, m/z 279.1699; Calcd for C15H23N2O3, m/z 279.1709 (M + H)⁺. Anal. (C₁₅H₂₂N₂O₃) C, H, N.

(1'S,4'R)-1-[4-(Hydroxymethyl)cyclopent-2-enyl]thymine (10). A mixture of compound 9 (50 mg, 0.18 mmol), CF₃CO₂H (8 mL), and H₂O (4 mL) was heated at 50 °C for 3 h. The solution was then concentrated to dryness and coevaporated with EtOH (2×5 mL). The residue was purified by silica gel column chromatography with 3% MeOH in CHCl₃ to give **10** (37 mg, 93%) as a white solid: mp 162–164 °C; [α]²⁴_D 71.66° (c 0.30, MeOH); UV (H₂O) λ_{max} 273.5 (ε 12757) (pH 2), 273.5 (ε 12893) (pH 7), 272.0 nm (e 9453) (pH 11); ¹H NMR (DMSOd₆) δ 7.34 (s, 1H, 6-H), 6.08 (m, 1H, 2'-H), 5.68 (m, 1H, 3'-H), 5.49 (m, 1H, 1'-H), 4.74 (t, J = 5.2 Hz, 1H, OH, D_2O exchangeable), 3.45 (m, 2H, 6'-Ha,b), 2.78 (m, 1H, 4'-H), 2.46 (m, 1H, 5'-Ha), 1.74 (s, 3H, CH₃), 1.33 (m, 1H, 5'-Hb); ¹³C NMR $(DMSO-d_6) \delta$ 163.05, 150.00, 138.31, 136.63, 128.96, 108.10, 62.55, 59.36, 46.32, 32.06, 11.31; HR-FAB MS Obsd, m/z223.1082; Calcd for $C_{11}H_{15}N_2O_3$, m/z 223.1083 (M + H)⁺. Anal. $(C_{11}H_{14}N_2O_3)$ C, H, N.

N-[[(1*S*,2*R*,3*S*,4*S*)-4-(*tert*-Butoxymethyl)-2,3-(isopropylidenedioxy)cyclopentyl]aminocarbonyl]-3-methoxy-2methyl-2-propenamide (12). To a solution of β -methoxy- α methacryloyl chloride (1.274 g, 9.46 mmol) in anhydrous benzene (13.2 mL) was added silver cyanate (3.04 g). The mixture was refluxed for 30 min and allowed to cool to room temperature. After the solid phase had settled, 6.33 mL of the supernatant solution which contained β -methoxy- α -methacryloyl isocyanate was added to a solution of amine **11** (820 mg) in dried DMF (16 mL) at -15 to -20 °C during 15 min. The reaction mixture was stirred at -15 °C for 2 h and then at room temperature for 8 h under nitrogen. The mixture was evaporated under reduced pressure, and the residue was purified by flash silica gel column chromatography with 5% MeOH in CHCl₃ to afford **12** (1.12 g, 88%) as a solid: mp 129–130 °C; $[\alpha]^{27}_{D}$ 22.52° (*c* 0.54, CHCl₃); ¹H NMR (CDCl₃) δ 7.30 (s, 1H, 6-H), 4.45 (m, 2H, 1'-H and 2'-H), 4.20 (m, 1H, 3'-H), 3.86 (s, 3H, OCH₃), 3.38 (m, 2H, 6'-H), 2.37 (m, 1H, 5'-H), 2.27 (m, 1H, 4'-H), 1.76 (s, 3H, 5-CH₃), 1.59 (s, 1H, NH), 1.57 (m, 1H, 5'-H), 1.49 (s, 3H, CH₃), 1.29 (s, 3H, CH₃), 1.17 (s, 9H, *tert*-butyl). Anal. (C₁₉H₃₂N₂O₆) C, H, N.

(1'*S*,2'*R*,3'*S*,4'*S*)-1-[4-(*tert*-Butoxymethyl)-2,3-(isopropylidenedioxy)cyclopentan-1-yl]thymine (14). Compound 12 (550 mg, 1.43 mmol) was dissolved in dioxane (4 mL), ethanol (4 mL), and aqueous ammonia (30%, 3 mL). The reaction mixture was heated at 80–100 °C in a sealed bomb for 10 h. The solution was evaporated to dryness. The residue was purified by flash silica gel column chromatography with 0.3% MeOH in CHCl₃ to give 14 (314 mg, 62%) as a solid: mp 154– 156 °C; [α]²⁵_D 35.53° (*c* 0.53, CHCl₃); UV (MeOH) λ_{max} 272.5 nm; ¹H NMR (CDCl₃) δ 7.08 (s, 1H, 6-H), 4.68 (m, 2H, 1'-H and 2'-H), 4.46 (m, *J* = 4.2, 6.0 Hz, 1H, 3'-H), 3.46 (m, 2H, 6'-H), 2.33 (m, 2H, 4'-H and 5'-H), 1.97 (m, 1H, 5'-H), 1.93 (s, 3H, 5-CH₃), 1.54 (s, 3H, CH₃), 1.30 (s, 3H, CH₃), 1.20 (s, 9H, *tert*-butyl). Anal. (C₁₈H₂₈N₂O₅) C, H, N.

N-[[(1*S*,2*R*,3*S*,4*S*)-4-(*tert*-Butoxymethyl)-2,3-(isopropylidenedioxy)cyclopentyl]aminocarbonyl]-3-methoxy-2propenamide (13). Conversion of 11 (1.6 g, 6.6 mmol) to 13 was accomplished using a procedure similar to that described for 12. The residue obtained was purified by silica gel column chromatography with 15% EtOAc in hexanes to give 13 (2.0 g, 82.1%) as a solid: mp 121–122 °C; $[\alpha]^{27}_{\rm D}$ 26.63° (*c* 0.57, CHCl₃); ¹H NMR (CDCl₃) δ 7.76 (br s, 1H, NH), 7.67 (d, *J* = 12.2 Hz, 1H, 6-H), 5.41 (d, *J* = 12.2 Hz, 1H, 5-H), 4.45 (m, 2H, 1'-H and 2'-H), 4.20 (m, 1H, 3'-H), 3.73 (s, 3H, OCH₃), 3.33– 3.42 (m, 2H, 6'-H), 2.36–2.43 (m, 1H, 5'-H), 2.28 (m, 1H, 4'-H), 1.57 (m, 1H, 5'-H), 1.46 (s, 3H, CH₃), 1.28 (s, 3H, CH₃), 1.17 (s, 9H, *tert*-butyl); ¹³C NMR (CDCl₃) δ 167.98, 163.06, 155.09, 111.47, 97.64, 86.46, 82.45, 72.80, 62.51, 57.48, 56.54, 45.08, 33.91, 27.38, 27.21, 24.85. Anal. (C₁₈H₃₀N₂O₆) C, H, N.

(1'*S*,2'*R*,3'*S*,4'*S*)-1-[4-(*tert*-Butoxymethyl)-2,3-(isopropylidenedioxy)cyclopentan-1-yl]uracil (15). A mixture of compound 13 (600 mg, 1.62 mmol), ethanol (10 mL), and aqueous ammonia (30%, 2.5 mL) was heated at 80–90 °C in a steel bomb for 24 h. After cooling, the solution was evaporated to dryness. The residue was purified by preparative TLC (3% MeOH in CHCl₃) to give 15 (493 mg, 90%) as an oil: $[\alpha]^{25}_{\rm D}$ 43.16° (*c* 0.67, CHCl₃); UV (MeOH) $\lambda_{\rm max}$ 266.5 nm; ¹H NMR (CDCl₃) δ 7.35 (d, *J* = 8.0 Hz, 1H, 6-H), 5.73 (d, *J* = 8.0 Hz, 1H, 6-H), 4.72 (m, 2H, 1'-H and 2'-H), 4.48 (m, 1H, 3'-H), 3.46 (m, 2H, 6'-H), 2.37 (m, 2H, 4'-H and 5'-H), 1.97 (m, 1H, 5'-H), 1.55 (s, 3H, CH₃), 1.30 (s, 3H, CH₃), 1.19 (s, 9H, *tert*-butyl); ¹³C NMR (CDCl₃) δ 167.98, 163.05, 155.09, 111.47, 97.64, 86.46, 82.45, 72.80, 62.51, 57.48, 56.54, 45.08, 33.91, 27.38, 27.21, 24.85. Anal. (C₁₇H₂₆N₂O₅·0.5H₂O) C, H, N.

(1'S,2'R,3'S,4'S)-1-[4-(tert-Butoxymethyl)-2,3-dihydroxycyclopentan-1-yl]thymine (16). A solution of compound 14 (362 mg, 1.03 mmol) in a mixture of concentrated HCl and MeOH (35.5 mL, concentrated HCl:MeOH = 1:70, v/v) was stirred at room temperature for 2.5 h. The reaction mixture was cooled in an ice bath and neutralized with NaHCO₃ solid. The mixture was concentrated to dryness. The residue was purified by silica gel column chromatography with 2-4%MeOH in CH₂Cl₂ to give **16** (258.5 mg, 80.6%) as a syrup: UV (MeOH) $\lambda_{\text{max}} 273.5$ nm; $[\alpha]^{24}_{\text{D}} 38.57^{\circ}$ (*c* 0.12, CHCl₃); ¹H NMR (DMSO- d_6) δ 11.20 (s, 1H, NH, D₂O exchangeable), 7.51 (s, 1H, 6-H), 4.82 (d, J = 6.5 Hz, 1H, OH, D₂O exchangeable), 4.63 (dd, 1H, J = 9.3, 18.5 Hz, 1'-H), 4.57 (d, J = 3.9 Hz, 1H, OH, D₂O exchangeable), 4.01 (m, 1H, 3'-H), 3.29 (m, 2H, 6'-Ha,b), 2.03-1.94 (m, 2H, 5'-Ha and 4'-H), 1.78 (s, 3H, CH₃), 1.24 (m, 1H, 5'-Hb), 1.14 (s, 9H, tert-butyl); HR-FAB MS Obsd, m/z 313.1766; Calcd for C₁₅H₂₅N₂O₅, m/z 313.1763 (M + H)⁺. Anal. (C15H24N2O5·0.2H2O) C, H, N.

(1'*S*,2'*R*,3'*S*,4'*S*)-1-[4-(*tert*-Butoxymethyl)-2,3-dihydroxycyclopentan-1-yl]uracil (17). A solution of compound 15 (3.2 g, 9.45 mmol) in a mixture of concentrated HCl and MeOH (341.2 mL, concentrated HCl:MeOH = 1:65, v/v) was stirred at room temperature for 3 h. The reaction mixture was cooled in an ice bath and neutralized with solid NaHCO₃. The mixture was concentrated to dryness. The residue was purified by silica gel column chromatography with 5–8% MeOH in CHCl₃ to give **17** (2.82 g, 100%) as a syrup: $[\alpha]^{25}_{D}$ 36.73° (*c* 0.65, CHCl₃); UV (MeOH) λ_{max} 267.5 nm; ¹H NMR (DMSO-*d*₆) δ 7.68 (d, *J* = 8.0 Hz, 1H, 6-H), 5.60 (d, *J* = 8.0 Hz, 1H, 5-H), 4.88 (d, *J* = 6.4 Hz, 1H, OH, D₂O exchangeable), 4.65 (m, 1H, 1'-H), 4.61 (d, *J* = 6.0 Hz, 1H, OH, D₂O exchangeable), 3.65 (m, 1H, 3'-H), 3.27 (m, 2H, 6'-Ha,b), 2.07–1.95 (m, 2H, 5'-Ha and 4'-H), 1.23 (m, 1H, 5'-Hb), 1.13 (s, 9H, *tert*-butyl); HR-FAB MS Obsd, *m*/*z* 299.1605; Calcd for C₁₄H₂₃N₂O₅, *m*/*z* 299.1607 (M + H)⁺.

(1'*S*,4'*R*)-1-[4-(*tert*-Butoxymethyl)cyclopent-2-enyl]uracil (20). Conversion of 17 (952 mg, 3.2 mmol) to 20 was accomplished using a procedure similar to that described for 9 (method b). The obtained residue was purified by silica gel column chromatography with 1% MeOH in CHCl₃ to give 20 (665 mg, 78.8%) as a solid: mp 107–110 °C; $[\alpha]^{26}_{D}$ 77.58° (*c* 0.99, CHCl₃); UV (MeOH) λ_{max} 267.5 nm; ¹H NMR (CDCl₃) δ 8.76 (br s, 1H, NH, D₂O exchangeable), 7.69 (d, *J* = 8.0 Hz, 1H, 5-H), 6.06 (m, 1H, 2'-H), 5.75 (m, 1H, 1'-H), 5.65 (d, *J* = 8.0 Hz, 1H, 6-H), 5.58 (m, 1H, 3'-H), 3.52 (dd, *J* = 9.0, 3.6 Hz, 1H, 6'-Ha), 3.34 (dd, *J* = 8.9, 4.1 Hz, 1H, 6'-Hb), 2.96 (m, 1H, 4'-H), 2.69 (dt, *J* = 14.4, 9.5, 9.5 Hz, 1H, 5'-Ha), 1.52 (m, 1H, 5'-Hb), 1.16 (s, 9H, *tert*-butyl); HR-FAB MS Obsd, *m*/*z* 265.1562; Calcd for C₁₄H₂₁N₂O₃, *m*/*z* 265.1552 (M + H)⁺. Anal. (C₁₄H₂₀N₂O₃) C, H, N.

(1'*S*,4'*R*)-1-[4-(Hydroxymethyl)cyclopent-2-enyl]uracil (21). Conversion of 20 (165 mg, 0.65 mmol) to 21 was accomplished using a procedure similar to that described for 10. The residue obtained was purified by silica gel column chromatography with 1–3% MeOH in CHCl₃ to give 21 (117 mg, 87%) as a white solid: mp 196–198 °C; $[\alpha]^{25}_{D}$ 104.68° (*c* 0.81, MeOH); UV (H₂O) λ_{max} 268.5 (ϵ 10540) (pH 2), 268.0 (ϵ 11562) (pH 7), 265.5 nm (ϵ 10019) (pH 11); ¹H NMR (DMSOd₆) δ 11.25 (br s, 1H, NH, D₂O exchangeable), 7.46 (d, *J* = 8.0 Hz, 1H, 5-H), 6.09 (m, 1H, 2'-H), 5.69 (m, 1H, 3'-H), 5.58 (d, *J* = 8.0 Hz, 1H, 6-H), 5.49 (m, 1H, 1'-H), 4.73 (t, *J* = 5.2 Hz, 1H, OH, D₂O exchangeable), 3.45 (m, 2H, 6'-Ha,b), 2.79 (m, 1H, 4'-H), 2.47 (m, 1H, 5'-Ha), 1.34 (m, 1H, 5'-Hb); HR-FAB MS Obsd, *m*/*z* 209.0926; Calcd for C₁₀H₁₃N₂O₃, *m*/*z* 209.0926 (M + H)⁺. Anal. (C₁₀H₁₂N₂O₃) C, H, N.

(1'S,4'R)-1-[4-(Hydroxymethyl)cyclopentan-1-yl]thymine (22). Compound 10 (40 mg, 0.45 mmol) in MeOH (10 mL) was hydrogenated at 15 psi in the presence of 10% Pd/C (30 mg) for 3 h at room temperature. The reaction mixture was filtered, and the filtrate was evaporated. The residue was purified by silica gel column chromatography with 2-3%MeOH in CHCl₃ to give **22** (36 mg, 89.2%) as a white solid: mp 172–174 °C; $[\alpha]^{28}_{D}$ 11.30° (*c* 0.18, MeOH); UV (H₂O) λ_{max} 274.0 (ε 10601) (pH 2), 273.5 (ε 10764) (pH 7), 273.0 nm (ϵ 6055) (pH 11); ¹H NMR (DMSO- d_6) δ 11.19 (br s, 1H, NH, D₂O exchangeable), 7.57 (s, 1H, 6-H), 4.72 (m, 1H, 1'H), 4.57 (t, 1H, OH, D₂O exchangeable), 3.36 (d, 2H, J = 6.3Hz, 6'-Ha,b), 2.06-1.33 (m, 7H, 2'-Ha,b, 3'-Ha,b, 4'-H, and 5'-Ha,b), 1.77 (s, 3H, CH₃); HR-FAB MS Obsd, m/z 225.1245; Calcd for C₁₁H₁₇N₂O₃, m/z 225.1239 (M + H)⁺. Anal. $(C_{11}H_{16}N_2O_3)$ C, H, N.

(1'*S*,4'*R*)-1-[4-(Hydroxymethyl)cyclopentan-1-yl]uracil (23). Conversion of 21 (72 mg, 0.45 mmol) to 23 was accomplished using a procedure similar to that described for 22. The residue obtained was purified by silica gel column chromatography with 1-3% MeOH in CHCl₃ to give 23 (65 mg, 90%) as a syrup: $[\alpha]^{28}{}_{\rm D}$ 21.28° (*c* 0.42, MeOH); UV (H₂O) $\lambda_{\rm max}$ 269.0 (ϵ 9949) (pH 2), 268.5 (ϵ 10085) (pH 7), 266.0 nm (ϵ 7646) (pH 11); ¹H NMR (DMSO-*d*₆) δ 11.22 (br s, 1H, NH, D₂O exchangeable), 7.70 (d, *J* = 8.0 Hz, 1H, 5-H), 5.57 (d, *J* = 8.0 Hz, 1H, 6-H), 4.73 (m, 1H, 1'H), 4.57 (t, 1H, OH, D₂O exchangeable), 3.38 (m, 2H, 6'-Ha,b), 2.08–1.31 (m, 7H, 2'-Ha,b, 3'-Ha,b, 4'-H, and 5'-Ha,b); HR-FAB MS Obsd, *m*/*z* 211.1081; Calcd for C₁₀H₁₅N₂O₃, *m*/*z* 211.1083 (M + H)⁺. Anal. (C₁₀H₁₄N₂ O₃·0.85H₂O) C, H, N.

(1'*S*,4'*R*)-1-[4-(*tert*-Butoxymethyl)cyclopent-2-enyl]cytosine (25). To a solution of compound 20 (400 mg, 1.51 mmol) in pyridine (23 mL) while stirring in a cold water bath was added dropwise 4-chlorophenyl dichlorophosphate (0.74 mL, 6.05 mmol), followed by the addition of 1,2,4-triazole (418 mg, 6 mmol). The mixture was stirred at room temperature for 24 h, and then the reaction mixture was evaporated in vacuo to dryness under 40 °C. The residue was dissolved in CH₂Cl₂ (25 mL), and the solution was washed with water (2 \times 50 mL), and saturated NaHCO₃ (50 mL). The organic layer was dried (MgSO₄), filtered, and concentrated to dryness to give crude 24 which was used for the next step without further purification. The crude 24 was stirred in a mixture of 30% aqueous ammonia (30 mL) and 1,4-dioxane (30 mL) at room temperature for 24 h. The solution was removed to dryness, and the residue was purified by silica gel column chromatography with 1-3% MeOH in CHCl₃ to give **25** (163 mg, 41%) as a white solid: mp >220 °C (dec); $[\alpha]^{28}_{D}$ 102.31° (*c* 0.50, CHCl₃); UV (MeOH) λ_{max} 276 nm; ¹H NMR (CDCl₃) δ 7.71 (d, J = 7.0 Hz, 1H, 5-H), 6.04 (m, 1H, 2'-H), 5.90 (m, 1H, 1'-H), 5.63 (d, J = 7.0 Hz, 1H, 6-H), 5.59 (m, 1H, 3'-H), 3.49 (dd, J = 3.9, 8.9 Hz, 1H, 6'-Ha), 3.31 (dd, J = 4.4, 8.8, Hz, 1H, 6'-Hb), 2.94 (m, 1H, 4'-H), 2.74 (dt, J = 14.4, 9.5, 9.4 Hz, 1H, 5'-Ha), 1.45 (dt, J = 14.3, 5.5, 5.1 Hz, 1H, 5'-Hb), 1.16 (s, 9H, tert-butyl); HR-FAB MS Obsd, *m*/*z* 264.1714; Calcd for C₁₄H₂₂N₃O₂, *m*/*z* 264.1712 $(M + H)^+$. Anal. $(C_{14}H_{21}N_3O_2)$ C, H, N.

(1'*S*,4'*R*)-1-[4-(Hydroxymethyl)cyclopent-2-enyl]cytosine (26). Conversion of 25 (136 mg, 0.52 mmol) to 26 was accomplished using a procedure similar to that described for 10. The obtained residue was purified by silica gel column chromatography with 10% MeOH in CHCl₃ to give 26 (101 mg, 94%) as a white solid: mp 150–153 °C; $[\alpha]^{28}_{D}$ 133.89° (*c* 0.44, MeOH); UV (H₂O) λ_{max} 284.5 (ϵ 10022) (pH 2), 274.5 (ϵ 7419) (pH 7), 274.5 nm (ϵ 7141) (pH 11); ¹H NMR (DMSO-*d*₆) δ 7.46 (d, *J* = 7.3 Hz, 1H, 5-H), 6.08 (m, 1H, 2'-H), 5.73 (d, *J* = 7.3 Hz, 1H, 6-H), 5.66 (m, 1H, 3'-H), 5.54 (m, 1H, 1'H), 4.70 (t, *J* = 5.0 Hz, 1H, OH, D₂O exchangeable), 4.12 (br s, 2H, NH₂, D₂O exchangeable), 3.38 (m, 2H, 6'-Ha,b), 2.77 (m, 1H, 4'-H), 2.47 (m, 1H, 5'-Ha), 1.25 (m, 1H, 5'-Hb); HR-FAB MS Obsd, *m*/*z* 208.1078; Calcd for C₁₀H₁₃N₃O₂, *m*/*z* 208.1086 (M + H)⁺.

(1'S,4'R)-1-[4-(Hydroxymethyl)cyclopentan-1-yl]cytosine (27). Compound 26 (52 mg, 0.25 mmol) in MeOH (15 mL) was hydrogenated at 15 psi in the presence of 10% Pd/C (40 mg) for 1 h at room temperature. The reaction mixture was filtered, and the filtrate was evaporated. The residue was purified by silica gel column chromatography with 8-10% MeOH in $CHCl_3$ to give **27** (45 mg, 85.6%) as a solid: mp 103– 105 °C; $[\alpha]^{28}_{D}$ 41.29° (*c* 0.22, MeOH); UV (H₂O) λ_{max} 285 (ϵ 12707) (pH 2), 275 (< 8529) (pH 7), 275.5 nm (< 9402) (pH 11); ¹H NMR (DMSO- d_6) δ 7.63 (d, J = 7.3 Hz, 1H, 6-H), 7.00 (br s, 2H, NH₂, D₂O exchangeable), 5.68 (d, J = 7.1 Hz, 1H, 5-H), 4.77 (m, 1H, 1'-H), 4.55 (t, J = 5.3 Hz, 1H, OH, D₂O exchangeable), 3.35 (m, 2H, 6'-Ha,b), 2.07-1.23 (m, 7H, 2'-Ha,b, 3'-Ha,b, 4'-H, and 5'-Ha,b); HR-FAB MS Obsd, m/z 210.1243; Calcd for $C_{10}H_{16}N_3O_2$, m/z 210.1243 (M + H)⁺. Anal. (C₁₀H₁₅N₃O₂•0.1CHCl₃) C, H, N.

(1'*S*,2'*R*,3'*S*,4'*S*)-9-[4-(*tert*-Butoxymethyl)-2,3-dihydroxycyclopentan-1-yl]-6-chloropurine (29). A solution of compound **28** (1.53 g, 4.01 mmol) in a mixture of concentrated HCl (2.62 mL) and MeOH (170 mL) was stirred at room temperature for 3 h. The reaction mixture was cooled in an ice–water bath and then neutralized with NaHCO₃ solid. The mixture was concentrated to dryness, and the residue was purified by silica gel column chromatography with 1-3% MeOH in CHCl₃ to give **29** (1.19 g, 86.8%) as a foam: $[\alpha]^{25}{}_{\rm D}$ 19.92° (*c* 0.67, CHCl₃); UV (MeOH) $\lambda_{\rm max}$ 263.5 nm; ¹H NMR (DMSO-*d*₆ + D₂O) δ 8.79 and 8.77 (two s, 2H, 2-H and 8-H), 4.85 (dd, *J* = 9.5, 18.7 Hz, 1H, 1'-H), 3.88 (dd, *J* = 5.0, 9.2 Hz, 1H, 2'-H), 3.80 (dd, *J* = 2.4, 4.9 Hz, 1H, 3'-H), 3.36 (m, 2H, 6'-Hab), 2.35 (dt, *J* = 8.9, 8.9, 13.0 Hz, 1H, 5-Ha), 2.08 (m, 1H, 4'-H), 1.78 (m, 1H, 5-Hb), 1.15 (s, 9H, *tert*-butyl); FAB MS *m/z* 341 (M + H)⁺.

(1'*S*,4'*R*)-9-[4-(*tert*-Butoxymethyl)cyclopent-2-enyl]-6chloropurine (31). Method a. A solution of diethyl azodicarboxylate (DEAD) (512 mg, 2.94 mmol) in dioxane (4.5 mL) was slowly added to a suspension of compound 8 (250 mg, 1.47 mmol), triphenylphosphine (770 mg, 2.94 mmol), and 6-chloropurine (454 mg, 2.94 mmol) in dry dioxane (15 mL). The mixture was stirred at room temperature for 10 h. The solvent was removed under reduced pressure, and the residue was purified by preparative TLC with 30% EtOAc in hexanes to give **31** (156 mg, 34.6%) as white solid.

Method b. A suspension of compound 29 (880 mg, 2.58 mmol) and pyridinium *p*-toluene-sulfonate (710 mg, 2.83) mmol) in trimethyl orthoformate (3 mL) was stirred at room temperature for 2 h. The reaction mixture was diluted with EtOAc (100 mL) and washed with saturated NaHCO₃ solution $(2 \times 15 \text{ mL})$ and brine (15 mL). The organic layer was dried (MgSO₄), filtered, and concentrated to dryness. The residue was coevaporated with benzene (2 \times 15 mL) and then treated with Ac₂O (14 mL). The mixture was heated at 120-130 °C for 12 h and then concentrated to dryness. The residue was purified by silica gel column chromatography with 15% EtOAc in hexanes to give 31 (429 mg, 54.2%) as a white solid: mp 111–114 °C; $[\alpha]^{22}_{D}$ 17.69° (c 1.21, CHCl₃); ¹H NMR (CDCl₃) δ 8.75 (s, 1H, 2-H), 8.45 (s, 1H, 8-H), 6.02 (m, 1H, 2'-H), 5.84 (m, 2H, 1'-H and 3'-H), 3.53 (dd, J = 4.2, 8.9 Hz, 1H, 6'-Ha), 3.39 (dd, J = 4.6, 8.9 Hz, 1H, 6'-Hb), 3.10 (m, 1H, 4'-H), 2.87 (dt, J = 14.3, 9.2, 9.2 Hz, 1H, 5'-Ha), 1.78 (m, 1H, 5'-Hb), 1.18 (s, 9H, tert-butyl); HR-FAB MS Obsd, m/z 307.1326; Calcd for $C_{15}H_{20}N_4ClO, m/z 307.1326 (M + H)^+$. Anal. ($C_{15}H_{19}N_4ClO$) C, H, N.

(1'S,4'R)-9-[4-(tert-Butoxymethyl)cyclopent-2-enyl]adenine (32). A solution of compound 31 (145 mg, 0.47 mmol) and saturated NH₃ in methanol (20 mL) was heated at 80-90 °C in a steel bomb for 20 h. After cooling, the solvent was removed under vacuum and the residue was purified by silica gel column chromatography with 0-1% MeOH in CHCl₃ to give compound 32 (112 mg, 82.5%) as a white solid: mp 182 C; UV (MeOH) λ_{max} 261 nm; $[\alpha]^{24}_{D}$ 19.86° (*c* 0.70, CHCl₃); ¹H NMR (CDCl₃) & 8.36 (s, 1H, 2-H), 8.04 (s, 1H, 8-H), 6.16 (m, 1H, 2'-H), 5.82 (m, 1H, 3'-H), 5.74 (m, 1H, 1'-H), 5.61 (br s, 2H, NH₂, D₂O exchangeable), 3.46 (dd, J = 4.8, 8.8 Hz, 1H, 6'-Ha), 3.35 (dd, J = 5.1, 8.8 Hz, 1H, 6'-Hb), 3.04 (m, 1H, 4'-H), 2.82 (dt, J = 14.1, 9.1, 9.1 Hz, 1H, 5'-Ha), 1.69 (m, 1H, 5'-Hb), 1.16 (s, 9H, tert-butyl); HR-FAB MS Obsd, m/z 288.1818; Calcd for $C_{15}H_{22}N_5O$, m/z 288.1824 (M + H)⁺. Anal. ($C_{15}H_{21}N_5O$) C. H. N.

(1'*S*,4'*R*)-9-[4-(*tert*-Butoxymethyl)cyclopent-2-enyl]-9*H*purine-6-thiol (33). A solution of compound 31 (165 mg, 0.54 mmol) and thiourea (52 mg, 0.67 mmol) in EtOH (17 mL) was refluxed for 1 h and cooled in an ice–water bath. The white precipitate formed was filtered and washed with EtOH (2 mL) to give a white solid of 33 (152 mg, 93%): mp 220 °C (dec); UV (MeOH) λ_{max} 325.0 nm; [α]²⁸_D –89.08° (*c* 0.21, CHCl₃); ¹H NMR (DMSO-*d*₆) δ 8.23 (s, 1H, 2-H), 8.05 (s, 1H, 8-H), 6.19 (m, 1H, 2'-H), 5.80 (m, 1H, 3'-H), 5.68 (m, 1H, 1'-H), 4.48 (dd, *J* = 8.7, 4.8 Hz, 1H, 6'-Ha), 3.38 (dd, *J* = 8.9, 5.0 Hz, 1H, 6'-Hb), 3.05 (m, 1H, 4'-H), 2.82 (dt, *J* = 14.1, 9.3, 9.2 Hz, 1H, 5'-Ha), 1.73 (m, 1H, 5'-Hb), 1.18 (s, 9H, *tert*-butyl); HR-FAB MS Obsd, *m*/*z* 305.1410; Calcd for C₁₅H₂₁N₄OS, *m*/*z* 305.1436 (M + H)⁺.

(1'S,4'R)-9-[4-(Hydroxymethyl)cyclopent-2-enyl]adenine (34). Conversion of 32 (50 mg, 0.18 mmol) to 34 was accomplished using a procedure similar to that described for 10. The residue obtained was purified by silica gel column chromatography with 3-5% MeOH in CHCl₃ to give 34 (34.5 mg, 86%) as a white solid: mp 189–192 °C; $[\alpha]^{24}$ 4.81° (*c* 0.52, MeOH); UV (H₂O) λ_{max} 260.5 (ε 14944) (pH 2), 261.5 (ε 16043) (pH 7), 261.5 nm (ϵ 15151) (pH 11); ¹H NMR (DMSO- d_6) δ 8.13 (s, 1H, 2-H), 8.04 (s, 1H, 8-H), 7.21 (br s, 2H, NH₂, D₂O exchangeable), 6.14 (m, 1H, 2'-H), 5.92 (m, 1H, 3'-H), 5.58 (m, 1H, 1'-H), 4.75 (t, J = 5.4 Hz, 1H, OH, D₂O exchangeable), 3.46 (m, 2H, 6'-Ha,b), 2.90 (m, 1H, 4'-H), 2.67 (dt, J = 13.7, 8.8, 8.6 Hz, 1H, 5'-Ha), 1.63 (m, 1H, 5'-Hb); ¹³C NMR (DMSO d_6) δ 154.80, 151.34, 151.31, 137.96, 137.44, 128.63, 117.80, 62.84, 58.12, 46.69, 33.29; HR-FAB MS Obsd, m/z 232.1181; Calcd for $C_{11}H_{14}N_5O$, m/z 232.1198 (M + H)⁺. Anal. ($C_{11}H_{13}N_5O$) C. H. N.

(1'S,4'R)-9-[4-(Hydroxymethyl)cyclopentan-1-yl]-9Hpurine-6-thiol (35). Conversion of 33 (143 mg, 0.47 mmol) to **35** was accomplished using a procedure similar to that described for **10**. The residue obtained was purified by silica gel column chromatography with 3% MeOH in CHCl₃ to give **35** (110 mg, 94%) as a solid: mp 235–238 °C; $[\alpha]^{28}_{\rm D}$ -41.61° (*c* 0.22, Pyr); UV (H₂O) $\lambda_{\rm max}$ 322.5 (ϵ 12953) (pH 2), 320.0 (ϵ 19421) (pH 7), 310.5 nm (ϵ 20659) (pH 11); ¹H NMR (DMSO-*d*₆) δ 13.72 (br s, 1H, NH, D₂O exchangeable), 8.20 (s, 1H, 2-H), 8.19 (s, 1H, 8-H), 6.17 (m, 1H, 2'-H), 5.92 (m, 1H, 3'-H), 5.58 (m, 1H, 1'-H), 4.74 (t, *J* = 5.3 Hz, 1H, OH, D₂O exchangeable), 3.45 (t, *J* = 5.6 Hz, 2H, 6'-Ha,b), 2.90 (m, 1H, 4'-H), 2.67 (dt, *J* = 13.8, 8.8, 8.7 Hz, 1H, 5'-Ha), 1.63 (dt, *J* = 13.8, 5.6, 5.6 Hz, 1H, 5'-Hb); HR-FAB MS Obsd, *m*/*z* 249.0821; Calcd for C₁₁H₁₃N₄O₂, *m*/*z* 249.0810 (M + H)⁺. Anal. (C₁₁H₁₂N₄OS) C, H, N.

(1'*S*,4'*R*)-9-[4-(Hydroxymethyl)cyclopentan-1-yl]adenine (36). Conversion of 34 (55 mg, 0.24 mmol) to 36 was accomplished using a procedure similar to that described for 22. The residue obtained was purified by silica gel column chromatography with 3–5% MeOH in CHCl₃ to give 36 (54 mg, 81%) as a white solid: mp 124–126 °C; $[\alpha]^{28}_{\rm D}$ –1.56° (*c* 0.29, MeOH); UV (H₂O) $\lambda_{\rm max}$ 260 (ϵ 15415) (pH 2), 261 (ϵ 16052) (pH 7), 261 nm (ϵ 166360) (pH 11); ¹H NMR (DMSO-d₆) δ 8.23 (s, 1H, 2-H), 8.12 (s, 1H, 8-H), 7.21 (br s, 2H, NH₂, D₂O exchangeable), 3.44 (m, 2H, 6'-Ha,b), 1.61–2.33 (m, 7H, 2'-Ha,b, 3'-Ha,b, 4'-H, and 5'-Ha,b); HR-FAB MS Obsd, *m*/*z* 234.1351; Calcd for C₁₁H₁₆N₅O, *m*/*z* 234.1355 (M + H)⁺. Anal. (C₁₁H₁₅N₅O+0.3MeOH) C, H, N.

(1'S,4'R)-9-[4-(Hydroxymethyl)cyclopent-2-enyl]hypoxanthine (38). A mixture of chloropurine analogue 31 (150 mg, 0.49 mmol), 2-mercaptoethanol (140 mg, 1.9 mmol), and NaOMe (105 mg, 1.95 mmol) in methanol (25 mL) was refluxed for 24 h. The mixture was cooled, neutralized with glacial AcOH, and concentrated under reduced pressure. The residue was purified by silica gel column chromatography with 3% MeOH in CHCl₃ to give **37** (140 mg, 99.3%) as a white solid which was used for the next step without further purification. Conversion of 37 (135 mg, 0.47 mmol) to 38 was accomplished using a procedure similar to that described for 10. The obtained residue was purified by silica gel column chromatography with 10% MeOH in CHCl₃ to give 38 (105 mg, 96.6%) as a white solid: mp 270–272 °C; $[\alpha]^{28}$ _D –3.83° (*c* 0.38, MeOH); UV (H₂O) λ_{max} 250.5 (ϵ 12128) (pH 2), 250.0 (ϵ 12873) (pH 7), 255.0 nm (ϵ 12095) (pH 11); ¹H NMR (DMSO- d_6) δ 12.29 (br s, 1H, NH, D₂O exchangeable), 8.04 (s, 1H, 2-H), 7.99 (s, 1H, 8-H), 6.15 (m, 1H, 2'-H), 5.91 (m, 1H, 3'-H), 5.56 (m, 1H, 1'-H), 4.73 (t, J = 5.2 Hz, 1H, OH, D₂O exchangeable), 3.45 (t, J= 5.7 Hz, 2H, 6'-Ha,b), 2.87 (m, 1H, 4'-H), 2.67 (dt, J = 13.7, 8.7, 8.7 Hz, 1H, 5'-Ha), 1.63 (dt, J = 13.7, 5.8, 5.8 Hz, 1H, 5'-Hb); HR-FAB MS Obsd, *m*/*z* 233.1047; Calcd for C₁₁H₁₃N₄O₂, m/z 233.1039 (M + H)⁺. Anal. (C₁₁H₁₂N₄O₂) C, H, N.

(1'*S*,4'*R*)-9-[4-(Hydroxymethyl)cyclopentan-1-yl]hypoxanthine (39). Conversion of **38** (66 mg, 0.28 mmol) to **39** was accomplished using a procedure similar to that described for **22**. The obtained residue was purified by silica gel column chromatography with 10% MeOH in CHCl₃ to give **39** (54 mg, 81%) as a white solid: mp 246–248 °C; $[\alpha]^{28}_{\rm D}$ – 1.04° (*c* 0.20, MeOH); UV (H₂O) $\lambda_{\rm max}$ 249.5 (ϵ 11108) (pH 2), 249.5 (ϵ 11999) (pH 7), 254.5 nm (ϵ 15312) (pH 11); ¹H NMR (DMSO-*d*₆) δ 12.26 (br s, 1H, NH, D₂O exchangeable), 8.18 (s, 1H, 2-H), 8.02 (s, 1H, 8-H), 4.78 (m, 1H, 1'-H), 4.63 (t, 1H, OH, D₂O exchangeable), 3.41 (m, 2H, 6'-Ha,b), 2.28–1.62 (m, 7H, 2'-Ha,b, 3'-Ha,b, 4'-H, and 5'-Ha,b); HR-FAB MS Obsd, *m*/*z* 235.1196; Calcd for C₁₁H₁₅N₄O₂, *m*/*z* 235.1195 (M + H)⁺. Anal. (C₁₁H₁₄N₄O₂·0.1H₂O) C, H, N.

(1'*S*,2'*R*,3'*S*,4'*S*)-9-[4-(*tert*-Butoxymethyl)-2,3-dihydroxycyclopentan-1-yl]guanine (42). A solution of compound 40 (880 g, 2.54 mmol), dimethyl formamide (10.3 mL), triethyl orthoformate (20.5 mL), and concentrated HCl (0.82 mL) was stirred at 0-5 °C for 8 h and then at room temperature for 12 h. The solvent was removed, and the residue was stirred with 50% HAc (47 mL) for 16 h. The mixture was concentrated to dryness, and the residue was purified by silica gel column chromatography with 1-2% MeOH in CHCl₃ to give

(1'S,2'R,3'S,4'S)-9-[4-(tert-butoxymethyl)-2,3-dihydroxycyclopentan-1-yl]-2-amino-6-chloropurine (41) (638 g, 70.5%) as a foam: UV (MeOH) λ_{max} 306.5 nm. A solution of the above 2-amino-6-chloropurine analogue 41 (275 mg, 0.77 mmol), 2-mercaptoethanol (300 mg, 3.84 mmol), NaOMe (325 mg, 6.0 mmol), and MeOH (40 mL) was refluxed for 18 h. The mixture was cooled and neutralized with glacial acetic acid, and the solvent was removed. The residue was purified by silica gel column chromatography with 8-10% MeOH in CHCl₃ to give **42** (150 mg, 58%) as a white solid: mp 237–240 °C; $[\alpha]^{25}_{D}$ 16.13° (c 0.48, MeOH); UV (MeOH) λ_{max} 254 nm; ¹H NMR (DMSO-d₆) δ 7.79 (s, 1H, 8-H), 6.42 (br s, 2H, NH₂), 4.93 (d, J = 5.9 Hz, 1H, OH, D₂O exchangeable), 4.60 (d, J = 3.3 Hz, 1H, OH, D₂O exchangeable), 4.53 (dd, J = 9.6, 18.4 Hz, 1H, 1'-H), 4.20 (m, 1H, 2'-H), 3.75 (m, 1H, 3'-H), 3.47 (m, 2H, 6'-Hab), 2.24 (m, 1H, 5-Ha), 2.00 (m, 1H, 4'-H), 1.37 (m, 1H, 5-Hb), 1.15 (s, 9H, tert-butyl); HR-FAB MS Obsd, m/z 338.1843; Calcd for $C_{15}H_{24}N_5O_4$, m/z 338.1828 (M + H)⁺

(1'*S*,4'*R*)-9-[4-(*tert*-Butoxymethyl)cyclopent-2-enyl]guanine (44). Conversion of 42 (140 mg, 0.41 mmol) to 44 was accomplished using a procedure similar to that described for 20. The residue obtained was heated with a mixture of aqueous ammonia (1 mL), methanol (6 mL), and H₂O (1 mL) at 65 °C for 1.25 h. The solvent was removed, and the residue was purified by silica gel column chromatography with 5% MeOH in CHCl₃ to give 44 (56 mg, 45%): UV (MeOH) λ_{max} 254 nm; ¹H NMR (CDCl₃) δ 12.08 (br s, 1H, NH), 7.68 (s, 1H, 8-H), 6.38 (br s, 2H, NH₂), 6.12 (m, 1H, 2'-H), 5.82 (m, 1H, 4'-H), 5.48 (m, 1H, 3'-H), 3.30-3.50 (m, 2H, 6'-H), 2.98 (m, 1H, 4'-H), 2.75 (dt, J = 8.8, 9.0, 14.0 Hz, 1H, 5'-Ha), 1.64 (m, 1H, 5'-Hb), 1.16 (s, 9H, *tert*-butyl); HR-FAB MS Obsd, *m*/*z* 304.1780; Calcd for C₁₅H₂₂N₅O₂, *m*/*z* 304.1774 (M + H)⁺.

(1'S,4'R)-9-[4-(Hydroxymethyl)cyclopent-2-enyl]guanine, L-Carbovir (45). Conversion of 44 (40 mg, 0.13 mmol) to 45 was accomplished using a procedure similar to that described for 10. The residue obtained was purified by silica gel column chromatography with 10% MeOH in CHCl3 to give **45** (27 mg, 84%) as a solid: mp 223–224 °C (decomp.) [lit²³ for D-(-) form, 210-220 °C (decomp.); lit⁷ for (±) form, 254-256 °C; lit²² for l-form, 265–270 °C]; $[\alpha]^{28}_{D}$ 59.03° (c 0.51, MeOH) [lit²⁵ for L-carbovir, $[\alpha]^{20}_D$ 60° (*c* 0.25, MeOH); lit²² for L-carbovir, $[\alpha]^{23}$ 61.1° (*c* 0.3, MeOH)]; UV (MeOH) λ_{max} 253.5 nm; ¹H NMR (DMSO- d_6) δ 10.35 (br s, 1H, NH, D₂O exchangeable), 7.58 (s, 1H, 8-H), 6.43 (br s, 2H, NH₂), 6.10 (m, 1H, 2'-H), 5.86 (m, 1H, 3'-H), 5.34 (m, 1H, 1'-H), 4.71 (t, J = 5.4 Hz, 1H, OH, D₂O exchangeable), 3.43 (t, *J* = 5.8 Hz, 2H, 6'-Ha,b), 2.85 (m, 1H, 4'-H), 2.58 (m, 1H, 5'-Ha), 1.56 (dt, J = 13.7, 5.8, 5.7 Hz, 1H, 5'-Hb); HR-FAB MS Obsd, m/z 248.1148; Calcd for $C_{11}H_{14}N_5O_2$, m/z 248.1147 (M + H)⁺.

Acknowledgment. This research was supported by the U.S. Public Health Research grants (AI 32351 and AI 33655) and the Department of Veterans Affairs. We thank Dr. Michael G. Bartlett for performing mass spectroscopy.

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JM9901327